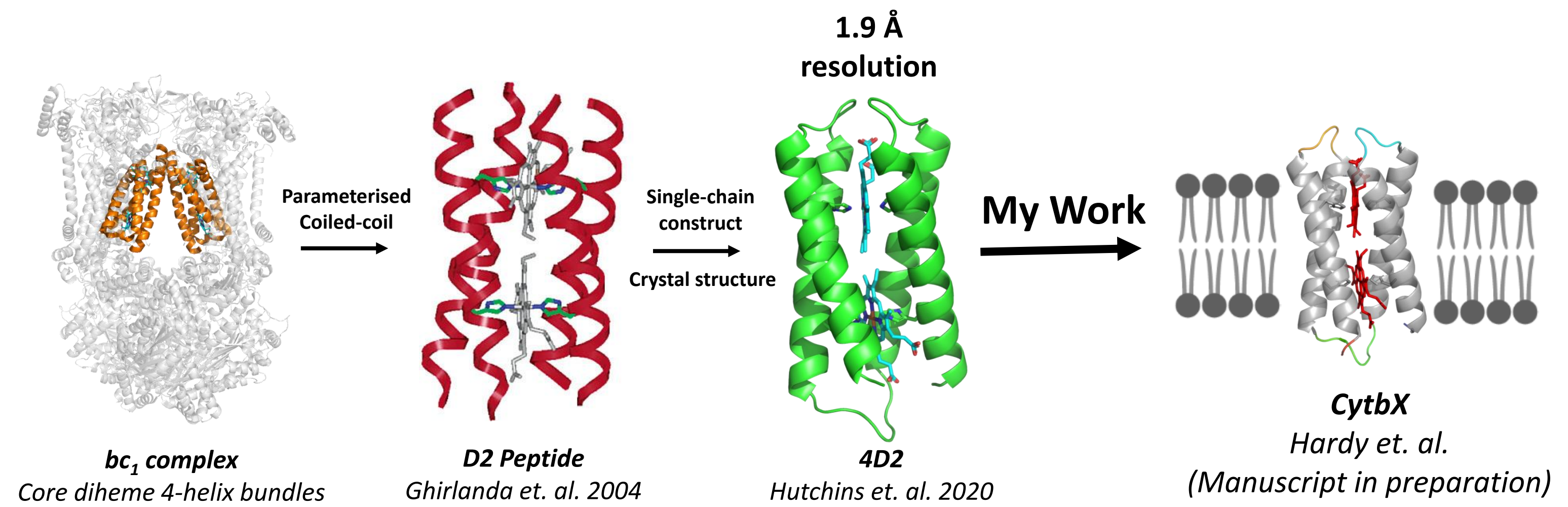




Background and Aims

- The aim of this work is to **convert** the water-soluble *de novo* design 4D2, a **parameterised maquette** of the bc_1 core **di-heme four-helix bundle**¹, to a **membrane protein** via computational design
- The purpose of the design is to be a **minimal module for transmembrane electron transfer**

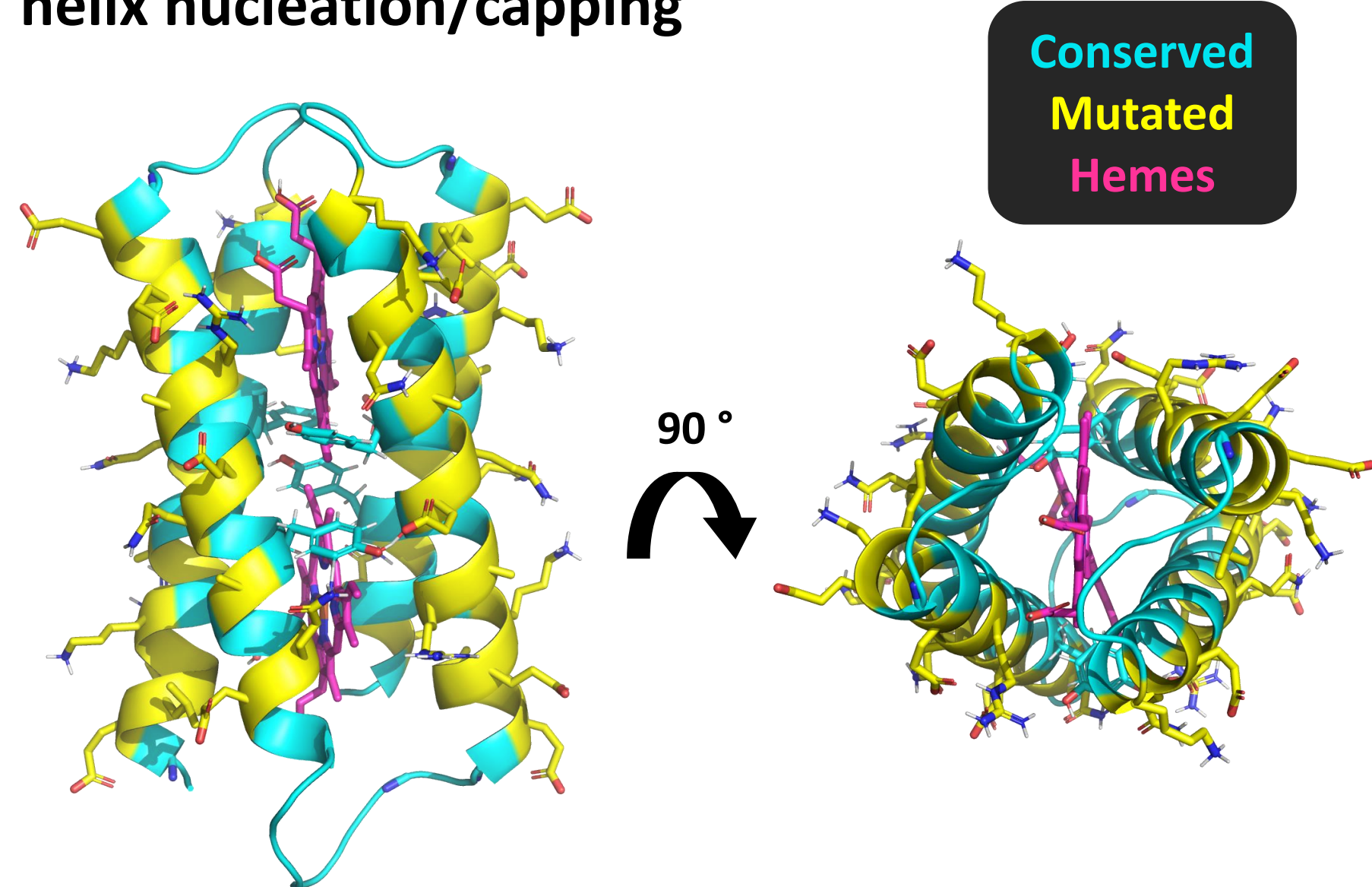


Computational Protein Design

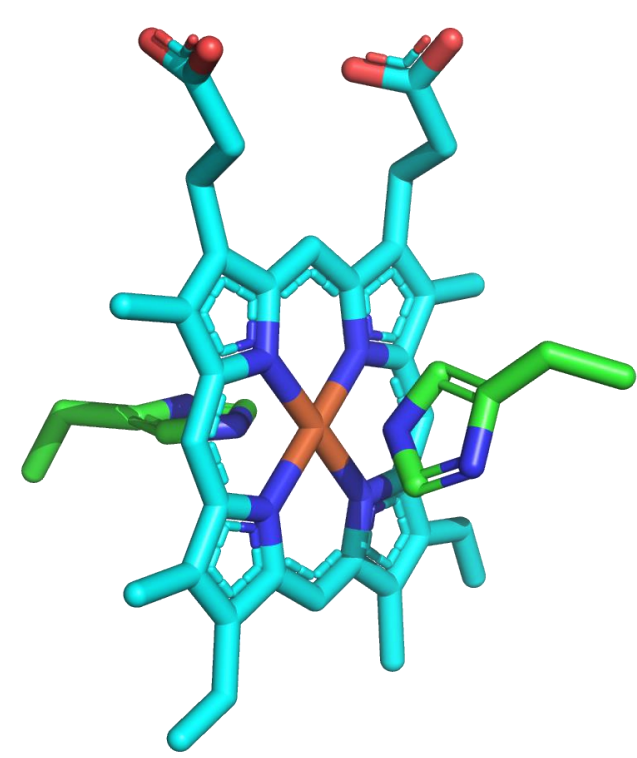
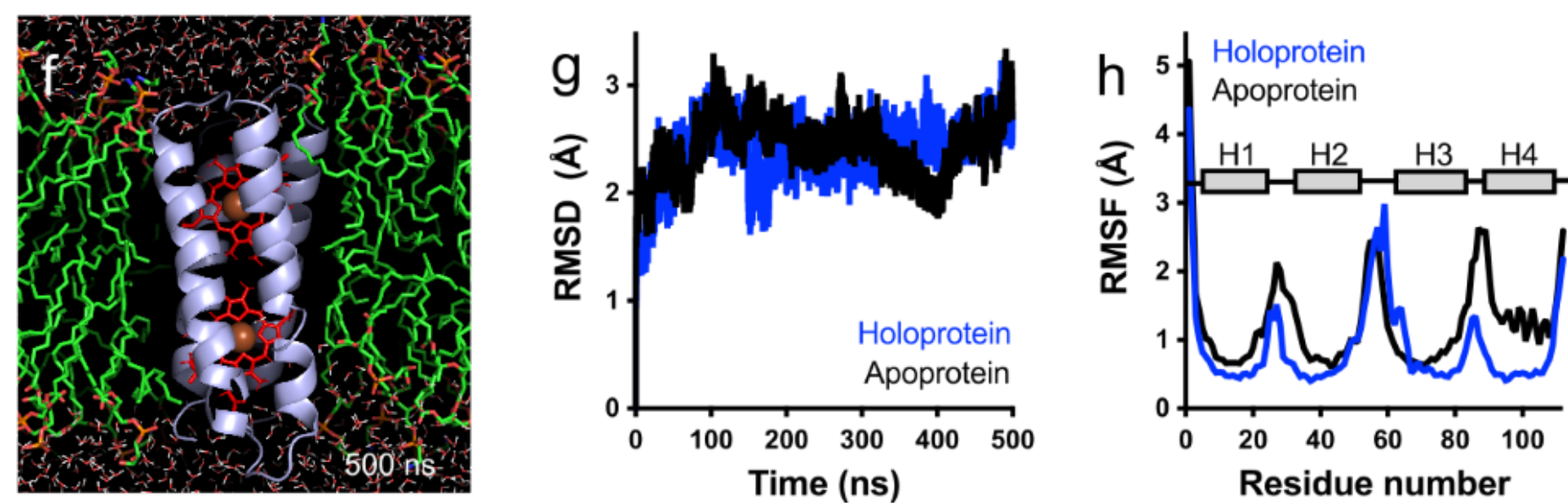
Rosetta was used to **mutate surface residues** of 4D2 to become lipophilic, whilst **preserving residues** responsible for **coiled-coil interactions**, **heme binding**, and **helix nucleation/capping**

Mutations were evaluated using the **franklin2019²** scorefunction

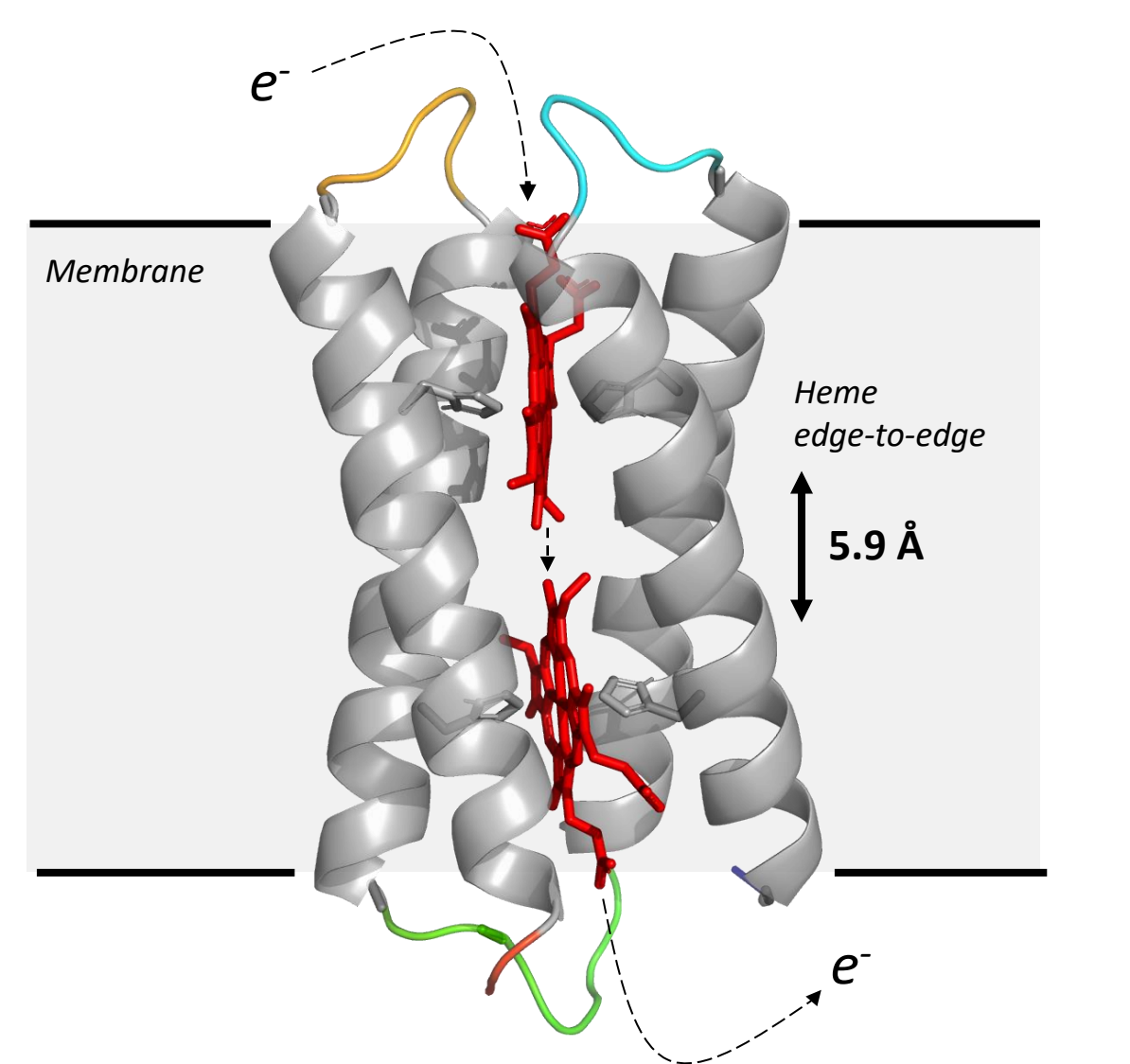
Designs were ranked based on total Rosetta score, quality of **core sidechain packing**, and number of **hydrogen bonds** to hemes



500 ns MD simulations showed CytbX is stable in a **3:1 DOPE:DOPG bilayer** in both the **apo** and **holo** states



Two b-type hemes bound by **bis-histidine coordination**, supplied by the **endogenous heme biosynthesis pathway** of *E. coli*



Protein Orientation: **Specified**
Electron transfer directionality: **Unspecified**

Anti-parallel four-helix bundle with **N_{in}-C_{in} topology** enforced by the **positive-inside rule** (charged loops)

Interfacial aromatic residues for anchoring at the **lipid head-group water boundary**

Leucine/isoleucine-rich hydrophobic surface

Knobs-into-holes packing motifs to specify **helix-helix interactions** in the membrane

Biophysical and Electrochemical Characterisation

UV-vis absorbance spectroscopy confirms binding of **heme b** by **bis-histidine coordination**

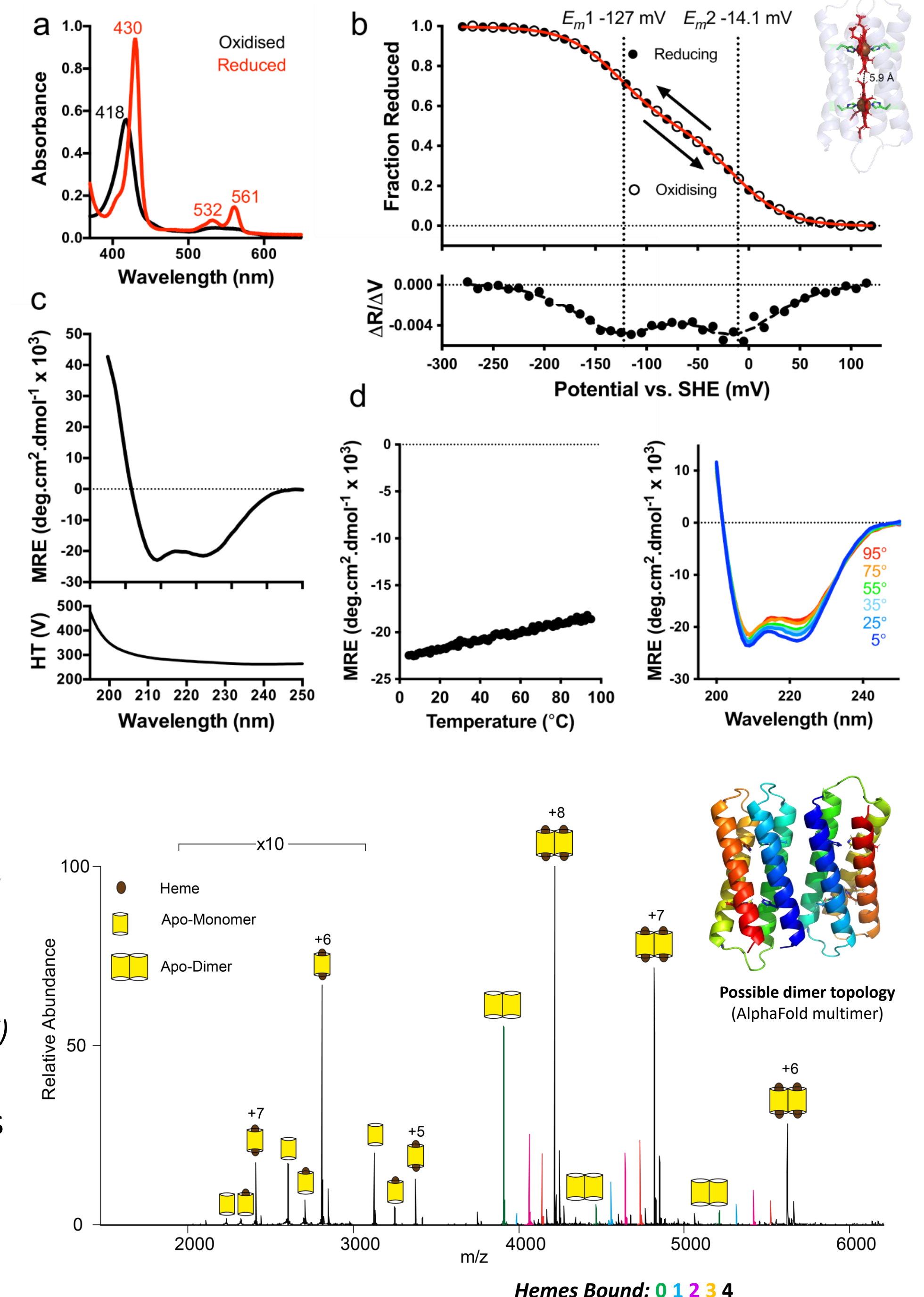
Two distinct **midpoint potentials** are observed, **split by about 110 mV**, indicating **electronic coupling** of **two hemes in close proximity**, as designed.

Resembles b_L and b_H of the bc₁ complex

Circular dichroism (CD) spectroscopy confirms that CytbX contains **alpha-helical** secondary structure, and reveals **exceptional thermal stability**

Native mass spectrometry (MS) of purified CytbX confirms binding of **two hemes per protein**

MS reveals that CytbX is purified as a **stable dimer**, reminiscent of the **dimeric interface of b subunits** in the **bc₁/b₆f complexes**

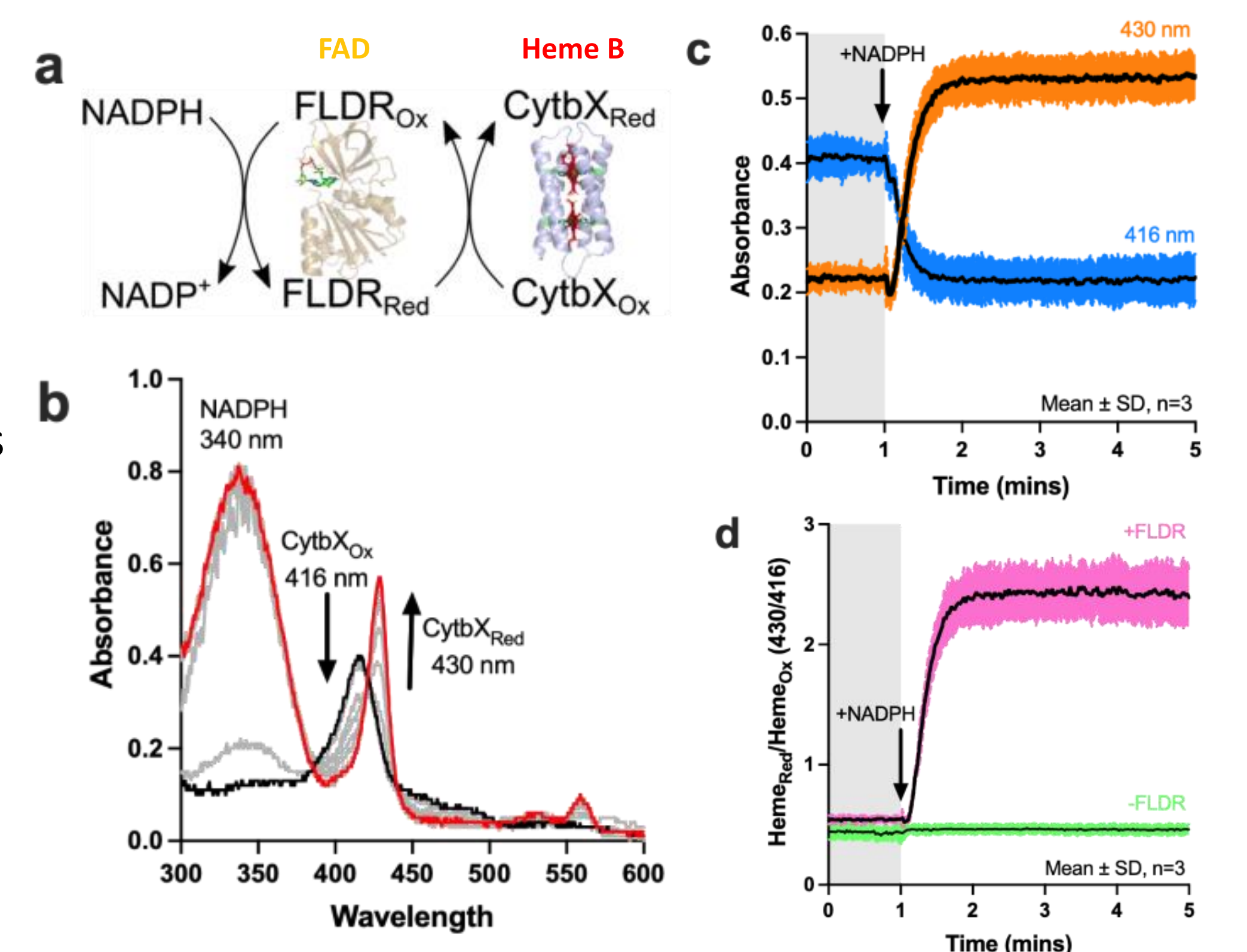


Minimal Electron Transport Chain

E. coli **flavodoxin reductase (FLDR)** can rapidly transfer electrons from **NADPH to the hemes** of CytbX (in micelles)

Oxidation state of the hemes is monitored **spectroscopically**

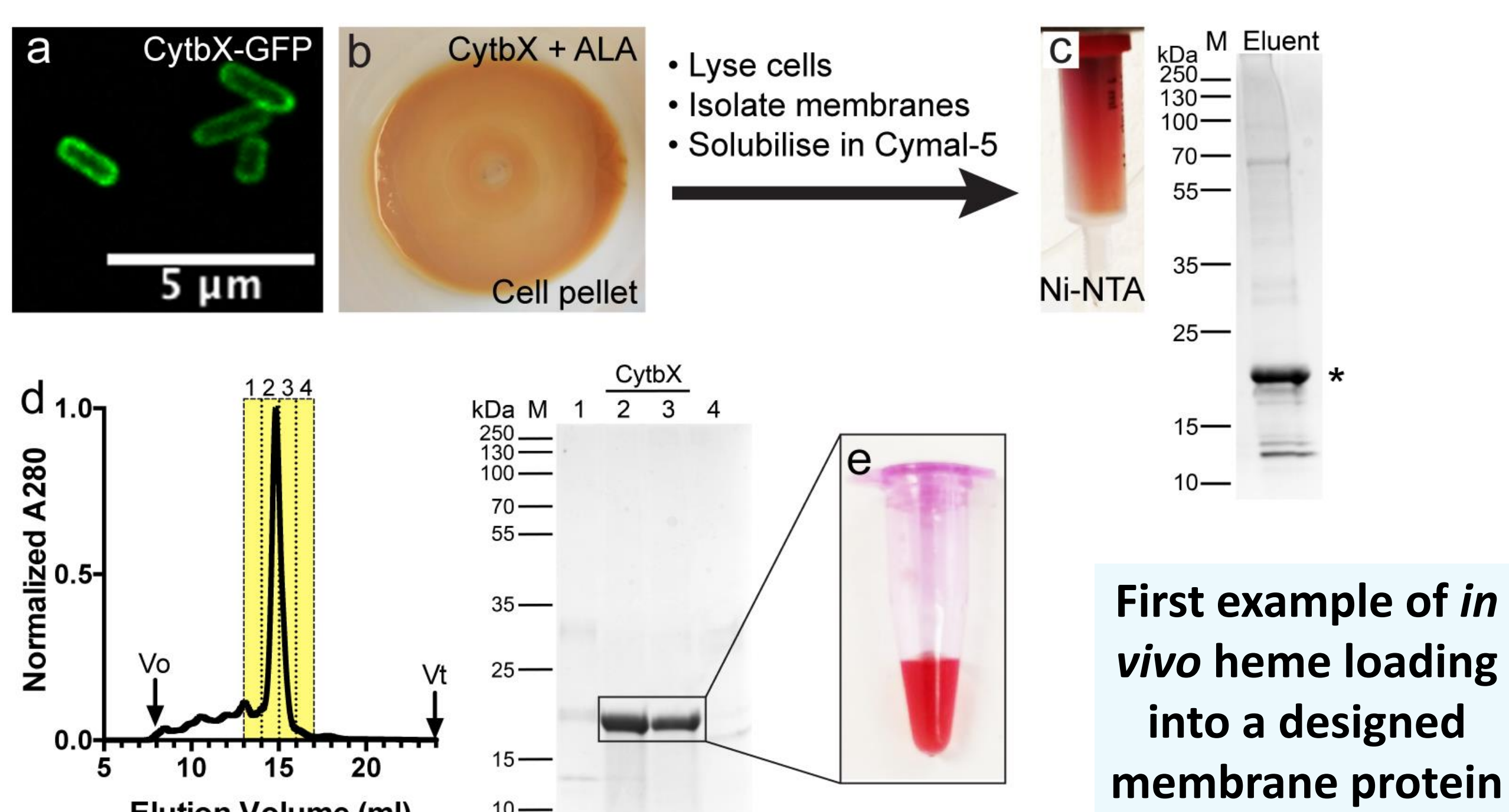
This demonstrates that the **membrane embedded hemes** are **accessible** to small diffusive redox partners



Recombinant Expression and Purification

CytbX is **recombinantly expressed** in **C43 (DE3) E. coli** and inserted into the plasma membrane by the natural translocation machinery.

Monodisperse, red-coloured CytbX can be **purified from E. coli membranes** using maltoside detergents
For purification methods see ref.³

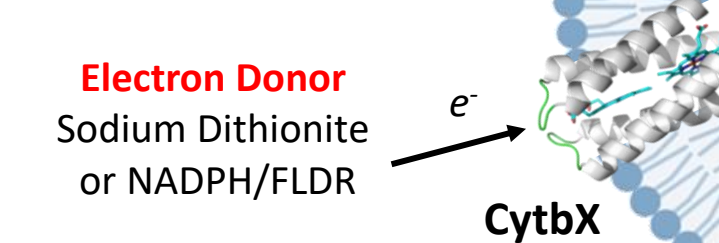


First example of *in vivo* heme loading into a designed membrane protein

Conclusions and Future Work

Conversion of 4D2 to a transmembrane protein was **successful**, and CytbX efficiently folds and binds the desired **cofactors *in vivo***.

Demonstrate transmembrane electron transfer in **proteoliposomes**



Alternative Cofactors

